Jaypee Institute of Information Technology

M.Sc. Microbiology

Semester I

Course Descriptions

<u>Detailed Syllabus</u> Lecture-wise Breakup

Course Code	15B1NBT832	Semester Od	ld	Semeste	er VII Session 2019-2020
		(specify Odd/Even)		Month July 19 to December 19	
Course Name	Biostatistics and Its applications				
Credits	4		Contact I	Hours	4
Faculty (Names)	Coordinator(s)	Shalini Mani			
	Teacher(s) (Alphabetically)	Shalini Mani			

COURSE O	COURSE OUTCOMES	
C430-3.1	Explain the various statistical methods to design a biological studies and data representation.	Understanding (Level 2)
C430-3.2	Apply different statistical methods and approaches to study the significance of a study.	Apply (Level 3)
C430-3.3	Examine the relationship between different parameters of a study.	Analyze (Level 4)
C430-3.4	Choose appropriate statistical methods, tools and resources including prediction, validation and evaluation of the biological studies.	Evaluate (Level 5)

Module No.	Title of the Module Module		No. of Lectures for the module	
1.	Introduction	Application and use of Biostatistics as a science, scope.	ics as a science, scope. 1	
2.	Study design in various fields of research	general principles of study design and its implications for valid inference	1	
3.	Sampling theory	Sampling scheme, simple/ systematic/ stratified/ cluster sampling, Sources of data collection	2	
4.	Data presentation	Graphical, tabular, Mathematical, finding the central tendency, measure of variations	3	
5.	Overview of different statistical methods used in the field of biological sciences.	Hypothesis testing, T-test, Chi square test, ANOVA, Sign Test, Wilcoxon Signed Rank Test, Wilcoxon Rank Sum Test, odds ratio, Binomial/normal/Poisson distribution of probabilities, determination of power of study and sample size calculation, regression analysis, correlation analysis,	13	
6.	Analysis of data source	Assess data sources and data quality for the purpose of selecting appropriate data for specific research questions	3	
7.	Selection of statistical methods	Identifying the appropriate statistical methods to be applied in a given research setting, applying the selected methods and analysis.	4	
8.	Application of Biostatistical	Designing various studies of medical/ health/ Microbial/Agricultural/Genetics/Pharamaceutical	7	

	analysis.	science related studies. Data analysis using different methods Result interpretation	
9.	Case studies	Based on various research studies and systematic reviews.	4
10.	SPSS, Stats at the bench	Introduction to SPSS, Entering data in SPSS editor. Solving the compatibility issues with different types of files. SPSS and working with descriptive statistics.	4
		Total number of Lectures	42

Evaluation Criteria	
Components	Maximum Marks
T1	20
T2	20
End Semester Examination	35
TA	25 (assignment, class test, quiz)
Total	100

Recommended Reading material: Author(s), Title, Edition, Publisher, Year of Publication etc. (Text books, Reference Books, Journals, Reports, Websites etc. in the IEEE format)
 Pranab Kumar Banerjee, Introduction to Biostatistics (4th Edition), S Chand and Company, 2015.
 Veer Bala Rastogi, Biostatistics (3rd Edition), Medtech, 2015
 S. Kartikeyan, R. M. Chaturvedi, R. M. Bhosale, Comprehensive textbook of biostatistics and research methodology(1st Edition), Bhalani Publishing House, 2016
 B Antonisamy Prasanna Premkumar Solomon Christopher, Principles and Practice of Biostatistics, Elsevier India, 2017
 Susan Holmes, Wolfgang Huber, Modern statistics for Modern Biology. Cambridge University Press, 2019

<u>Detailed Syllabus</u> Lecture-wise Breakup

2000010 1120 2100010					
Course Code	19M21BT113	Semester Odd		Semester M.Sc. Microbiology I	
				Session	2019 -2020
				Month	from July-December
Course Name	Biomolecu	iles			
Credits	4		Contact I	Hours	4
Faculty (Names)	Coordinator(s)	Dr. Reema Ga	abrani		

Teacher(s) Dr. Priyadarshini, Dr. Reema Gabrani	Faculty (Names)	Coordinator(s)	Dr. Reema Gabrani
(Alphabetically)		Teacher(s) (Alphabetically)	Dr. Priyadarshini, Dr. Reema Gabrani

COURSE	OUTCOMES	COGNITIVE LEVELS
C113.1	Explain the biomolecules structure and function	Understand Level (C2)
C113.2	Analyze bioenergetics and metabolic pathways for physiological and pathological conditions	Analyze Level (C4)
C113.3	Apply the concepts of enzymes, hormones and signaling	Applying Level (C3)
C113.4	Illustrate the basics in genomics and proteomics	Understand Level (C2)

Module No.	Title of the Module	Topics in the Module	No. of Lect ures for the mod ule
1.	Carbohydrates and Bioenergetics	Chemical composition and bonding; Carbohydrates: Classification, basic chemical structure; General reactions of the functional groups; Physiological significance; Metabolism of carbohydrate: Glycolysis, TCA, gluconeogenesis, PPP, ATP role; Respiratory chain and oxidative phosphorylation	11
2.	Lipids	Classification, structure and function of major lipid subclasses; chylomicrons, LDL, HDL, and VLDL; Pathological changes in lipid levels. Formation of micelles, monolayers, bilayer, liposomes; biosynthesis of fatty acids and ketogenesis	7
3.	Proteins	Amino acids: Classification, Properties, Protein Structure: primary, secondary, tertiary and quaternary structure; separation techniques; Enzymes: kinetics, functions; biosynthesis of non-essential amino acids and catabolism of protein and amino acids in born errors of metabolism.	7
4.	Nucleotides	Nucleic acid structure, Nucleotides and nucleosides; metabolism of purines and pyrimidines	6
5.	Hormones	Characteristics of hormones/ signalling molecules; function, signal transduction	6
6.	Introduction to Genomics and proteomics	DNA sequence analysis methods; gene disease association; Introduction and scope of proteomics	5

		Total number of Lectures 42
Evaluation Criteria		
Components	Maximum Marks	
T1	20	
T2	20	
End Semester Examination	35	
TA	25 (Presentation, Assignments)	
Total	100	

Ioui	100			
Recommended Reading material: Author(s), Title, Edition, Publisher, Year of Publication etc. (Text books, Reference Books, Journals, Reports, Websites etc. in the IEEE format)				
1.	JM Berg, L Stryer, J Tymoczko, G Gatto, "Biochemistry", 9th Ed. San Francisco, 2019 WH Freeman			
2.	B Gomberts, "Signal transduction", Academic Press, 2009 Harper			
3.	RK Murray, DK Granner, VW Rodwell, "Harper's Illustrated Biochemistry", 27 th Ed. McGraw-Hill Lange 2006			
4.	D Voet and JG Voet, "Biochemistry" 4 th Ed. Wiley 2010			
5.	DL Nelson and MM Cox, "Lehninger Principles of Biochemistry", 7 th Ed. WH Freeman 2017			

Detailed Syllabus Lecture-wise Breakup

Course Code	19M21BT111	Semester: Odd		Semeste	emester: I Session: 2019-2020	
				Month f	from: July to December	
Course Name	Microbial Physiology and Diversity					
Credits	4		Contact I	Hours	3-1-0	

Faculty (Names)	Coordinator(s)	Dr. Smriti Gaur
	Teacher(s) (Alphabetically)	Dr. Garima Mathur, Dr. Smriti Gaur

Course Outcomes:
At the completion of the course, students will be able to,

Sl. No.	DESCRIPTION	COGNITIVE LEVEL (BLOOM'S TAXONOMY)
C110.1	Classify the Diversity amongst archae, eubacteria and other microorganisms	Understanding level (Level 2)
C110.2	Demonstrate ecological diversity, habitat interaction and microbial relationship.	Understanding level (Level 2)
C110.3	Identify microbial nutritional, growth requirements and associated physiological mechanisms.	Applying level (Level 3)
C110.4	Analyze the different modes of metabolism in microorganisms.	Analyzing level(Level 4)

Module No.	Title of the Module	Topics in the Module	No. of Lectures for the module		
1.	Microbial taxonomy and evolution of diversity	taxonomy and molecular characteristics, Phylogenetic trees evolution of			
2.	The Archae (Extremophiles and their diversity)	Introduction to Archaeal Taxonomy and Metabolism, Phylum Crenarchaeota: Habitat and energy metabolism, cold dwelling microbes (artic and antartic regions), hyperthermophiles. Phylum Euryarchaeota: extremely halophilicarchea, taxonomy and physiology of halophilicarchea. Methanogens – diversity and physiology. Thermoplasmatales—thermoplasma, Hyperthermophilic euryarcheota: Thermococcales and Methanopyrus.	4		
3.	Gram negative and positive	Diversity, characteristic features and significance : Spirochaetes - aerobic / microaerophilic motile, helical /	5		

eubacteria	vibriod - non motile gram negative curved bacteria - gram negative and positive rod and cocci - gram negative straight, curved & helical rods - sulfur reducing bacteria - rickettisias and chlamydias – mycoplasmas - endosymbionts.	
	Mycobacteria – Nocardioformis. Anoxygenic phototrophic bacteria – oxygenic photosynthetic bacteria – aerobic chemolithotrophic bacteria – budding and appendaged bacteria – sheathed bacteria – non photosynthetic bacteria - Myxobacteria – archeobacteria.	
4. Diversity of other microorganisms	Distribution, importance, structure and characteristics of the fungal divisions, slime molds, the algal divisions, protozoans, general properties of viruses, their structures and classification, bacteriophages	7
5. Microbial Diversity of various habitats	Microorganisms in nature ecosystem, Ecological groups of Microorganisms, Microbial population interactions, Human-Microbe Interactions, The soil habitat, Water as a Microbial Habitat, Microflora of air, Microflora of foodstuff	5
6. Microbial nutrition and growth 7. Bacterial photosynthesis	Nutritional requirements of Microorganisms- Autotrophs, Heterotrophs, Chemotrophs, Copiotrophs and Oligotrophs. Transport Mechanisms - Diffusion- Facilitated Diffusion, Active transport- Group translocation. Different phases of growth - Growth curve - Generation time - Factors influencing microbial growth - Temperature, pH, Pressure, Salt concentration, Nutrients - synchronous growth and continuous cultivation. Diauxic growth, Sporulation - Endospore formation in bacteria. Chemotherapeutic agents as growth inhibitors Photosynthetic microorganisms, photosynthetic pigments, and generation of reducing power by	5
	cyclic and non-cyclic photophosphorylation, electron transport chain in photosynthetic bacteria. Carbon dioxide fixation pathways.	
8. Bacterial Respiration	Bacterial aerobic respiration, components of electron transport chain, free energy changes and electron transport, oxidative phosphorylation and theories of ATP formation, inhibition of electron transport chain. Electron transport chain in some heterotrophic and chemolithotrophic bacteria. Bacterial anaerobic respiration: Introduction. Nitrate, carbonate and sulfate as electron acceptors. Electron transport chains in some anaerobic bacteria. Catalase, super oxide dismutase, mechanism of oxygen toxicity.	5
9. Bacterial Chemolithotrophy	Physiological groups of chemolithotrophs, ammonia oxidation by members of Genus Nitroso group, nitrite oxidation by Nitro group of genera. Oxidation of molecular hydrogen by <i>Hydrogenomonas</i> species. Ferrous and sulfur/sulfide oxidation by <i>Thiobacillus</i> species.	4
1	Total number of Lectures	42

Eval	uation Criteria		
Com	ponents	Maximum Marks	
T1	•	20	
T2		20	
End	Semester Examination	35	
TA		25	
Tota	l	100	
Reco	mmended Reading mater	ial: Author(s), Title, Edition, Publisher, Year of Publication etc. (Text	
book	s, Reference Books, Journ	nals, Reports, Websites etc. in the IEEE format)	
1.	Microbial Diversity by Co	lwd , D. 1999, Academic Press.	
2.	Prescott L M, J P Harley a	nd D A Klein (2005). Microbiology. Sixth edition, International	
	edition, McGraw Hill.		
3.	Advances in Applied Mici	obiology. Vol. 10. Edited by Wayne W. Umbreit and D.	
•	Pearlman. Academic Press	.	
4.	Brocks Biology of Microorganisms. 8th Edition. (International Edition - 1997) by		
	Michael T. Madigan, John M. Martinko. Jack Parker. Prentice Hall Internation Inc.		
5.		mentals and Applications by. Ronald M. Atlas and Richard	
	Bartha. 2nd and 4th Edition	n. The Benjamin Cummins Publication Co. Inc.	
6.	¥ •	gy and biochemistry of prokaryotes. Oxford university press. 4th	
	edition (2011).		

<u>Detailed Syllabus</u> Lecture-Wise Breakup

Course Code	19M21BT115	Semester: Odd	Semester: 1 st	Session: 2019-2020	
			Month from: Ju	ly to December	
Course Name	Microbial Genetics & Molecular Biology				
Credits	3-1-0-4		Contact Hours	4	

COURSE	OUTCOMES: Upon completion of the course, students will be able to	COGNITI VE LEVELS
CO112.1	• Explain fundamental principles of molecular biology and technological advances in the field	Understa nding Level (C2)
CO112.2	Apply knowledge of microbial genome architecture and gene regulation	Apply Level (C3)
CO112.3	• Analyse various methods of gene transfer and extrachromosomal inheritance	Analysis Level (C4)
CO112.4	Interpret different aspects of DNA mutations, DNA repair, Linkage & Mapping	Understa nding Level (C2)

Faculty	Coordinator(s)	1. Prof. Krishna Sundari					
(Names)	Teacher(s)						
	(Alphabetically)	2. Dr. Vibha Gupta					
Module No.	Subtitle of the Module	Topics in the module	No. of Lectures for the module				
1.	The nature of	ature of Discovery of DNA and experimental evidence, The					
	Genetic material	structure of DNA and RNA; Melting of DNA,					
		Superhelicity, Genome architecture, Chromatin					
		arrangement, nucleosome formation, C value					
		paradox, central dogma					
2.	DNA replication	DNA replication mechanism, enzymes involved and	06				
	and repair	models of DNA replication, DNA methylation,					
		inhibitors of DNA replication, DNA damage and					
		repair: Molecular basis of spontaneous and induced					
		mutations, types of mutation, Ames test, DNA					
		repair pathways - excision, mismatch,					
		photoreactivation, Double Strand Break Repair					

3.	DNA	Transcription machinery - various transcription	07
	transcription	enzymes and cofactors, initiation, elongation and	
	•	termination, enhancer sequences and control of	
		transcription, Structure and function of RNA	
		polymerase, Post-transcriptional processes: RNA	
		processing, Capping and polyadenylation, rRNA and	
		tRNA processing, RNA Editing; RNAi and	
		miRNAs, Antisense RNA	
4.	DNA translation	The genetic code and protein structure, Mechanisms	06
		of translation - initiation complex, ribosomes and	
		tRNA, factors, elongation and termination, in vitro	
		translation systems, polycistronic/monocistronic	
		synthesis, inhibitors of translation, stringent	
		response in bacteria, Post-translational processes:	
		Protein modification, folding, chaperones,	
		transportation; protein degradation	
5.	Methods of gene	Transformation - natural transformation systems,	04
	transfer in	mechanism, chemical-mediated and electro-	
	Bacteria	transformation; Conjugation - nature of donor	
		strains and compatibility, interrupted mating and	
		temporal mapping, F plasmid, Hfr transfer,	
		horizontal gene transfer	
6.	Plasmids &	Plasmid types, detection, replication, partitioning,	04
	Movable genetic	copy-number control, properties of some known	
	elements	plasmids, Extrachromosomal inheritance	
7.	Genetic control	Operons, lac system, trp system for negative &	04
	mechanism in	positive gene regulation, lambda phage, complex	
•	prokaryotes	operons	0.0
8.	Viral genome &	Introduction to viral genetics, viral life cycles and	03
	Methods of gene	phage replication, Transduction - Generalized and	
	transfer in	specialized transduction; gene mapping by	
0	Viruses	specialized transduction	02
9.	Linkage and gene	Recombination (homo and heterologous), linkage	03
	Mapping	symbolism, single and double cross overs, linkage	
10	Tankmalasissi	maps, genetic analysis	02
10.	Technological	Recombination as a molecular biology tool,	03
	advances	Genetically modified organisms (GMOs) and	
		applications Total number of Leatures	12
		Total number of Lectures	† <i>L</i>
Evaluati	ion Criteria	<u>"</u>	
Compor		Maximum Marks	
со тро г Г1		20	
Г2		20	
	nester Examination	35	
ТА		25	
Total		100	

Recommended Reading material: Author(s), Title, Edition, Publisher, Year of Publication etc. (Text books, Reference Books, Journals, Reports, Websites etc. in the IEEE format)

1	Lewin's Genes XII by Jocelyn E. Krebs, Elliott S. Goldstein and Stephen T. Kilpatrick	
1.	Jones and Bartlett Publishers, Sudbury, Massachusetts, 2018.	
2	Molecular Biology of the Gene by J.D. Watson, T.A. Baker, S.P. Bell, A. Gann, M. Levin,	
2.	R. Losick, 7th edition, Benjamin Cummings, San Francisco, USA, 2013.	
2	Molecular Biology of the Cell by B. Alberts, A. Johnson, J. Lewis, M. Raff, K. Roberts, P.	
Walter, 6th edition, Garland Science, New York and London, 2017.		
4	Lehninger Principles of Biochemistry Seventh Edition – David L. Nelson; Michael M.	
4.	Cox, 2017	
_	An Introduction to Genetic Analysis by Suzuki DT, Griffiths AJF, Miller JH and Lewontin	
5.	RC, WH freeman and Company, New York	

<u>Detailed Syllabus</u> Lab-wise Breakup

Course Code	19M25BT111	Semester Odd		Semeste	er I Session 2019 -2020
		(specify Odd/I	Even)	Month i	from July-December
Course Name	Microbiology Lab-I				
Credits	4 Contact		Hours	8	

Faculty (Names)	Coordinator(s)	Dr. Ashwani Mathur
	Teacher(s) (Alphabetically)	Dr. Ashwani Mathur, Dr. Indira P. Sarethy, Dr. Rachana, Prof. Neeraj Wadhwa, Dr. Shalini Mani, Prof. Sudha Srivastava, Prof. Sujata Mohanty, Prof. Vibha Rani

COURSE	OUTCOMES	COGNITIVE LEVELS
CO1	Understand various culture media, their applications and methods of sterilization	Understand (Level C2)
CO2	Apply standard microbiological techniques for isolation, culturing and enumeration of microorganisms	Apply (Level C3)
CO3	Make use of different methods for microbial identification and characterization	Apply (Level C3)
CO4	Compare methods of DNA isolation from microorganisms	Analyze (Level C4)

Module No.	Title of the Module	List of Experiments	Hours
1.	Isolation of microorganisms from different sources	Media preparation & sterilization – Bacteria; Media preparation & sterilization – fungi; Preparation of agar plants and slants; Culturing microorganisms on agar media by streaking / stab / point inoculation; Serial dilution of microbial culture; Estimation of microbial growth by colony counting	Week 1 – Week 3
2.	Characterization of Microorganisms	Microbial diversity – characterization of bacteria & fungi; IMVIC Test; Computational tool for strain identification	Week 4 – Week 6
3.	Microbial Growth	Effect of substrate / culture conditions on microbial growth; To study diauxic growth in bacteria; Data presentation & Analysis	Week 7 – Week 9
4.	Molecular Biology	Isolation of DNA from bacteria; Isolation of bacteria from fungi; Agarose Gel Electrophoresis	Week 10– Week 12
		Total	12

Evaluation Criteria

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Components	Maximum Marks
Mid-Term Viva	20
Day-to-Day (Lab record,	
attendance, performance)	60
Final Viva	20
Total	100

Recommended Reading material: Author(s), Title, Edition, Publisher, Year of Publication etc. (Text books, Reference Books, Journals, Reports, Websites etc. in the IEEE format)

1.	https://microbeonline.com/imvic-tests-principle-procedure-and-results/
2.	Vashist Hemraj, Sharma Diksha, Gupta Avneet (2013), A review on commonly used biochemical test for bacteria Innovare Journal of Life Science, Vol 1: Issue 1, 1-7
3.	Manual of Microbiology: Tools and Techniques- Kanika Sharma ISBN 10: 8180520889 / ISBN 13: 9788180520884

<u>Detailed Syllabus</u> Lecture-wise Breakup

Course Code	19M21HS111	Semester: Odd		Semester: 2019 -2020	
				Month:	July-Dec 2019
Course Name	Presentation and Con	nmunication Ski	lls		
Credits	Credits 2		Contact I	Iours	2-0-0

Faculty (Names)	Coordinator(s)	Dr. Parineeta Singh
	Teacher(s) (Alphabetically)	Dr. Parineeta Singh

COURSE OUTCOMES		COGNITIVE LEVELS
C101.1	Develop an in-depth understanding and appreciate the subtle aspects of English as a communication tool.	Understand(C2)
C101.2	Assess the communication challenges of a diverse, global marketplace	Analyze (C4)
C101.3	Create & Compose different forms of Professional writing	Create (C6)
C101.4	Evaluate the effectiveness of sample Presentations	Evaluate (C5)
C101.5	Apply the acquired skills in delivering effective presentations	Apply (C3)

Module No.	Title of the Module	Topics in the Module	No. of Lectures for the module
1.	Communication Process, Grammar, and Vocabulary	 Communication: Definition, Model, Channel, Goals Process of Communication: Linear Concept, Shannon-Weaver Model, the Two-Way Process Communication Traits: Communication Apprehension, Style, Argumentativeness and Verbal Aggressiveness Grammar: denotative and connotative words, subject-verb agreement Techniques of Vocabulary Building 	5
2.	Intercultural Communication	 Recognizing cultural diversity: variations in a diverse world Developing Cultural Intelligence: High-Context Cultures and Low-Context Cultures Time as a cultural factor: Monochronic and Polychronic Time Challenges of Intercultural Communication Developing Cultural Competency and Guidelines for Adapting. 	5
3.	Business Etiquettes, and Presentation Skills	 Ekman's classification of communicative movements Face Facts, Positive Gestures, Negative Gestures, Lateral Gestures Preparing and Delivering a Presentation Using Audio-Visual Aids: Presentation Support 	5

4.	Communication for Conflict Management	 Sample Presentations: Steve Jobs, Three Stories of my Life (Stanford University Commencement Address, 2005) Dr. Shashi Tharoor, Britain does owe India reparations (Oxford Union Debate) Negotiation, Mediation, and Conciliation Stages in the Negotiation Process Strategies of Conciliation Solving Deadlocks Reaching an Agreement 	5
5.	Communication for Employment	 Guidelines for writing a Resume, Types of Resumes Interviews: Purpose and Types. Interviews: Preparation, Process, Common Mistakes to Avoid. Group Discussion: Stages (Forming, Storming, Norming, Performing, Adjourning) Formal/Informal Group Dynamics 	5
6.	Technical Communication	 Characteristics of a Report Types of Report 5 W's and 1 H of a Report Structure, Format, Parts of a Report Referencing, and Documentation 	5
		Total number of Lectures	30
Evalua	tion Criteria		
	onents rm Examination (Presenta mester Examination	Maximum Marks tion) 30 40 30(Assignment/ Viva) 100	

	Recommended Reading material: Author(s), Title, Edition, Publisher, Year of Publication etc. (Text books, Reference Books, Journals, Reports, Websites etc. in the IEEE format)		
1	C.L.Bovee, J.V.Thill, Roshan Lal Raina, <i>Business Communication Today</i> , 13 th Ed, Pearson Education, 2017.		
2	R.C. Sharma and Krishna Mohan, <i>Business Correspondence and Report Writing</i> , Mc Graw Hill Education, 2016.		
3	Meenakshi Raman and Sangeeta Sharma, <i>Technical Communication: Principles and Practice</i> , Oxford University Press, 2015.		
4	Anna Koneru, Professional Communication, Mc Graw Hill Education Pvt Ltd., 2017.		
5	Murli Krishna, Communication Skills for Engineers, Pearson, 2014.		
6	Meenu Dudeja, Communication Skills for Professionals, Satya Prakashan, 2017.		
7	Barun K. Mitra, <i>Personality Development and Soft Skills</i> , Oxford University Press, 2012.		